

Available online at www.sciencedirect.com



PHARMACOLOGY BIOCHEMISTRY ^{AND} BEHAVIOR

Pharmacology, Biochemistry and Behavior 81 (2005) 131-138

www.elsevier.com/locate/pharmbiochembeh

The influence of dextromethorphan on morphine analgesia in Swiss Webster mice is sex-specific

Judith E. Grisel^{a,*}, Stephani Allen^a, Kumar V.S. Nemmani^b, Jon R. Fee^c, Richard Carliss^d

^aDepartment of Psychology, Furman University, 3300 Poinsett Highway, Greenville, SC 29613, United States

^bDepartment of Psychology and Department of Psychology and Centre for Research on Pain, McGill University, Montreal, QC, Canada H3A 1B1

^cDepartment of Psychology at The University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, United States

^dEndo Pharmaceuticals, Inc., Chadds Ford, PA 19317 and Department of Clinical Laboratory Sciences, University of South Alabama Mobile,

AL 36604, United States

Received 17 September 2004; received in revised form 3 March 2005; accepted 4 March 2005 Available online 12 April 2005

Abstract

NMDA (*N*-methyl-D-aspartate) antagonists are known to enhance the analgesic effects of opioids. However, virtually, all studies of this phenomenon have been done using male subjects. Here, the noncompetitive NMDA receptor antagonist dextromethorphan (DEX) was tested over a range of doses (10–200 µg intracerebroventricularly [i.c.v.]) in male and female Swiss Webster mice in combination with 5 mg/kg intraperitoneal (i.p.) morphine. Males exhibited enhanced morphine analgesia following either 100 or 200 µg DEX, but there was no evidence of DEX-mediated potentiation in females at any dose. Instead, DEX attenuated morphine analgesia in females. We also evaluated the effect of 100 µg i.c.v. DEX with different doses of morphine (1, 5 and 10 mg/kg). Again, DEX significantly enhanced morphine analgesia in male mice and attenuated it in females. Next, ovariectomized (OVX) female mice were compared to males following 5 mg/kg i.p. morphine analgesia was not enhanced by DEX in sham-treated females and OVX mice with estradiol treatment (5 µg i.p. once daily for 7 days) also did not show DEX enhancement. These experiments demonstrate that the ability of NMDA receptor antagonists to potentiate morphine analgesia is modulated by an estrogen-sensitive mechanism and suggest that sex differences may play a critical role toward a more general understanding of the potentiation of opioid-induced analgesia through NMDA receptor antagonists.

Keywords: Morphine; NMDA [N-methyl-D-aspartate]; Dextromethorphan; Sex; Mice; Analgesia

Morphine has remained the drug of choice for the treatment of moderate to severe pain for well over a century. However, tolerance to morphine or other opiate drugs can be profound and debilitating, and detract markedly from the intended therapeutic benefits. Tolerance, defined as a decrease in the potency of a drug following repeated administration (Cox, 1990), can occur during a single administration of an opioid (Kornetsky and Bain, 1968). In order to enhance the therapeutic efficacy of opioids such as morphine, researchers have sought ways to minimize

* Corresponding author. Tel.: +1 864 294 3218.

E-mail address: judy.grisel@furman.edu (J.E. Grisel).

tolerance and other adverse effects (see Bhargava, 1994 for review).

NMDA receptors are thought to play a role in the adaptation to opioids. NMDA antagonists attenuate the development of tolerance to morphine (Trujillo and Akil, 1991; Tiseo and Inturrisisi, 1993; Lufty et al., 1999) and there is a large body of research demonstrating that blockade of NMDA receptors potentiates opiate analgesic efficacy over a range of assays in a variety of species (e.g., Chapman and Dickenson, 1992; Advokat and Rhein, 1995; Mao et al., 1996; Plesan et al., 1999; Wen et al., 2004). For example, enhancement of opiate analgesia by NMDA receptor blockade has been reported in various strains of male mice (Bernardi et al., 1996; Bhargava, 1997; Lufty et al., 1999;

Redwine and Trujillo, 2003), rats (Bespalov et al., 1998; Plesan et al., 1999; Christensen et al., 1998, 1999; Carlezon et al., 2000; Nishiyama, 2000; Kozela et al., 2001; Bulka et al., 2002; Laulin et al., 2002) and non-human primates (Allen and Dystra, 2001).

Despite what appears to be ample evidence for NMDA mediation of opioid analgesia, when NMDA receptor antagonists have been used to either enhance the analgesic effects of opiates or to attenuate tolerance, in the vast majority of studies, only male subjects have been used. This is noteworthy because there are known sex differences in pain thresholds and in the pharmacological effects of morphine and NMDA antagonists (e.g., Kepler et al., 1991; Mogil et al., 1993; Kavaliers and Galea, 1995; Cicero et al., 1996; Boyer et al., 1998; Kest et al., 1999; Sarton et al., 2000). Indeed, we recently published a broad survey of the effects of several NMDA antagonists on morphine analgesia and evaluated both male and female mice (Nemmani et al., 2004). In this study, we reported that sex, along with the site of antagonism, morphine dose and time after injection, all significantly influenced morphine response. Others have found similar results: Holtman et al. (2003) showed that sex influenced NMDA receptor-mediated effects on morphine analgesia in rats. It is also worth noting that not all studies, even using males, find unequivocal potentiation of morphine effects following NMDA antagonism (see Kozela et al., 2001 or Redwine and Trujillo, 2003 for reviews).

In order to further study the sex-dependency of opiate analgesia by NMDA antagonists, we compared the effect of dextromethorphan, a noncompetitive NMDA receptor antagonist, combined with morphine in male, female and ovariectomized female mice.

1. Materials and methods

1.1. Animals

Swiss Webster mice of both sexes were bred in-house from stock obtained from Taconic Farms (Germantown, NY) and maintained 2–4 per Thoren caging in a temperature $(21\pm2$ °C) and light (12 h reverse light/dark cycle; lights off at 0700 h) controlled colony room. Food and water were freely available at all times and corncob bedding was changed twice a week. Between 7 and 12 weeks of age, naïve mice were taken from the colony room in their home cages to a procedural room and allowed to habituate to the experimental room for at least 30 min before testing. All experiments were approved by the Furman University animal care and use committee and in accordance with NIH guidelines.

1.2. Nociceptive assay

Pain sensitivity was measured by determining the latency to reflexive withdrawal of the tail (TW) following immersion in warm water bath (Ben-Bassat et al., 1959). Animals were lightly restrained in a cloth container (voluntarily entered in all cases) and the distal half of the tail was immersed in 49 $^{\circ}$ C water. The latency to reflex withdrawal was measured using a stopwatch to the nearest 0.1 s by an experienced observer blind to drug condition. To improve accuracy, two TW latency determinations, separated by approximately 10 s, were made and averaged at each time point. A cut-off response latency was set at 15 s to prevent tissue damage.

1.3. Drug administration

Morphine sulfate (Sigma-Aldrich, St. Louis, MO) was dissolved in 0.9% saline and delivered intraperitoneally (i.p.) using a 26 g 3/8'' needle. Dextromethorphan (DEX; Sigma-Aldrich) was also dissolved in saline but with 5% acetic acid. DEX (or vehicle, saline with 5% acetic acid) was delivered intracerebroventricularly (i.c.v.). in order to insure that equal amounts of DEX were present in the central nervous system of male and female mice. I.c.v. injections were always in a volume of 2.5 µl/mouse and were made directly into the left lateral ventricle using the method of Laursen and Belknap (1986) under isofluorane anesthesia. Isofluorane is a safe, short-acting, inhalant anesthetic with no reported interactions with morphine, DEX or gonadal steroids.

1.4. Group designations and testing procedures

Data included in this study represent three separate experiments. In the first, we evaluated the effect of various doses of DEX (10, 100 or 200 µg) on analgesia resulting from 5 mg/kg morphine. We assessed TW latencies in male and female mice, delivered i.c.v. DEX or vehicle immediately followed by i.p. injection of morphine, and then reassessed TW latencies at 30, 60, 90 and 120 min. For the second experiment, we kept the dose of DEX constant at 100 μ g/mouse, but varied morphine doses (0, 1, 5 or 10 mg/ kg). We chose this middle dose of DEX to minimize the likelihood of either "floor" or "ceiling" effects in our twotailed analyses. Administration and testing procedures were the same as in the first experiment except that post-injection TW latencies were determined at 30, 60 and 120 min. In the last experiment, we evaluated the effect of 100 µg DEX in combination with 5 mg/kg morphine (again at 30, 60, 90 and 120 min post-injection) and we also included female subjects that varied with respect to hormone status (see below). There were 8-15 subjects per experimental group.

1.5. Surgical procedure and hormone replacement

Female mice in the last experiment involving gonadal hormones received either ovariectomy (OVX) or sham ovariectomy (SHAM) via dorsal incision under ketamine/ xylazine anesthesia as described in Mogil et al. (1993). Males were similarly anesthetized, but no surgical manipulation occurred. Animals were allowed 10–14 days between surgery

and testing in order to be sure recoveries were complete and gonadal hormones were depleted in OVX animals. On the first of 2 test days, half of each group was administered i.p. morphine+i.c.v. DEX (100 µg) and the other half received morphine+vehicle. Beginning 2-4 days after this assessment, ovariectomized mice were given daily intraperitoneal (i.p.) injections of 5 µg estradiol (Sigma-Aldrich) in 0.1 ml sesame oil for 7 days. Males and sham-treated females were administered equivolume vehicle (sesame oil) throughout this period. On the following day, all animals were re-tested for antinociception to morphine, in a crossover design following i.c.v. injection (subjects receiving saline during the first test got DEX and vice versa). Although ketamine is also an NMDA receptor antagonist, and thus could interact with subsequent DEX administration, the long intervening period (at least 10 days) decreases this possibility, but more importantly, because all mice received equal anesthetic treatment, this is especially unlikely to account for any sex differences.

1.6. Statistics

Raw TW data were analyzed using two-way (drug and sex) repeated measure analysis of variance (ANOVA). In

order to determine which groups were different from their controls, within-sex (single factor) ANOVAs were conducted and pair wise comparisons were evaluated using Dunnett's two-sided post-hoc test (SYSTAT 10.2). Raw TW data were also converted to antinociceptive area under the time×TW latency curve for each animal (AUC, min*s), analyzed by two-factor ANOVA, then further evaluated in the same manner as above. In all cases, the criterion for statistical significance was set at p < 0.05.

2. Results

The results of the first two experiments indicate that DEX influenced morphine analgesia differently in male and female mice. While there were no differences between the sexes in morphine-induced antinociception, DEX potentiated morphine analgesia in males, but attenuated it in females (indicated by a significant interaction between sex and drug condition on TW latencies: $F_{(3,46)}=8.90, p < 0.01$). Analyzed and presented are the mean TW latencies for each group as well as the geometric areas under the TW latency curves in order to depict both the time course (duration) and overall effects of the drug treatment in each group.



Fig. 1. The top panels of this figure depict the antinociceptive responses of female (A) or male (B) Swiss Webster mice to intracerebroventicular (i.c.v.) administration of either 10, 100 or 200 μ g dextromethorphan (DEX) followed immediately by intraperitoneal (i.p.) administration of 5 mg/kg morphine. For males given 100 and 200 μ g DEX, statistically significant differences in TW latencies compared to saline alone occurred at 90 and 120 min (*p <0.05, Dunnett's *t*). The lower panels summarize the same data collapsed for time in order to compare the areas under the analgesic curve (AUC) as a function of different doses of DEX in females (C) or males (D). DEX produced a dose-dependent attenuation of the AUCs in females, while a dose-dependent enhancement of the AUCs was produced in males. In both cases at 200 μ g DEX, AUCs were statistically different from saline (*p <0.05, Dunnett's *t*).

The top panels of Fig. 1 show the time course for the effects of different doses of DEX on morphine analgesia in both females (Fig. 1A) and males (Fig. 1B). Overall, there was more analgesia in males, due to potentiation and prolongation of morphine effects by DEX. Fig. 1C and D show the area under the analgesic curves (AUC) for each dose of DEX. In both males and females, there was a dose-dependent effect of DEX in which DEX induced an attenuation of analgesia in females at 200 µg i.c.v. and an enhancement of morphine analgesia in males at this same dose (Dunnett's test p < 0.05 in both cases).

In a [2 (sex)×4 (drug dose)] analysis, a main effect of sex ($F_{(1,46)}$ =10.93, p<0.01) was obtained supporting the observation that male mice were more analgesic overall. In addition, DEX changed the shape of the analgesia curve in both sexes as it altered the early effect of morphine in females and prolonged the effect of morphine in males. Thus, there was an interaction between time and sex ($F_{(4,184)}$ =2.6, p<0.05) and an interaction between time and drug condition ($F_{(12,184)}$ =2.24, p<0.05) as well as a triple interaction between time, sex and drug indicating that the effect of DEX on the analgesic curve was dependent upon sex. These results are depicted again in the lower panel, showing the AUC data and borne out by analysis: a main effect of sex ($F_{(1,46)}$ =8.37, p<0.05), and an interaction between sex and drug ($F_{(3,46)}$ =7.70,

p < 0.01). The potentiation seen in males following either of the higher two doses was still present 120 min after morphine administration (Dunnett's post-hoc test, p < 0.05 in both cases).

In the second experiment, testing the effect of DEX combined with different doses of morphine, DEX again affected morphine analgesia differently in males and females (Fig. 2). A repeated measure ANOVA indicated significant effects of drug ($F_{(5,92)}$ =5.23, p < 0.01), sex ($F_{(1,92)}$ =5.96, p < 0.05) and their interaction ($F_{(5,92)} = 3.39, p < 0.01$). There was also a time×drug interaction ($F_{(15,276)}$ =4.62, p<0.01), and again, a triple interaction between time, drug and sex $(F_{(15,276)}=1.85, p<0.05)$ reflecting the fact that DEX prolonged morphine analgesia in male mice (Fig. 2B). In males given 5 or 10 mg/kg of morphine in combination with DEX, analgesia lasted at least 2 h. This was not the case in males without DEX treatment, or in females receiving DEX (Fig. 2A), as these groups were all back to baseline nociceptive values at this time point. Using Dunnett's test to compare each of the drug treatment groups to a control group receiving neither morphine nor DEX (S-S), DEX was found to enhance morphine-induced analgesia in males only at 10 mg/kg. Moreover, DEX was found to obviate the analgesia seen in female mice receiving either 5 or 10 mg/kg DEX as neither of these doses produced significant analgesia when combined with 100 µg DEX. When combined with 1



Fig. 2. Panels A and B illustrate the effect for different doses of parental morphine when DEX was co-administered i.c.v. at a single dose of $100 \mu g$. DEX was given in combination with 1, 5 or 10 mg/kg i.p. morphine. DEX enhanced and prolonged morphine antinociception in males and attenuated it in females. The lower panels, showing the area under the analgesic curves (AUC), summarize these differential effects of DEX in males and females.

mg/kg morphine, there was no effect of DEX on morphineinduced analgesia in either sex. AUC data are shown in Fig. 2C (female) and D (males).

In the third experiment, ovariectomy reversed and estrogen replacement in ovariectomized mice restoredthe female-specific phenotype. Again, DEX potentiation of morphine analgesia was evident in males, as well as in ovariectomized female mice, but not in other groups (Fig. 3). ANOVA of the raw TW data for Experiment 3 (see Fig. 3A), analyzed in a 2 (drug condition; saline vs. 100 µg DEX × 4 (sex/hormone status) design showed a significant difference between the drug conditions $(F_{(1,130)}=12.45,$ p < 0.01) and a significant effect of sex $(F_{(3,130)} = 11.86,$ p < 0.01). Again, the interaction was significant, indicating that the effect of drug was dependent upon sex $(F_{(3,130)}=7.43, p < 0.01)$. Within subjects, there was an effect of time $(F_{(4,520)}=279.60, p<0.01)$ and a significant interaction between time and drug ($F_{(4,520)}=3.38$, p<0.05) as well as a triple interaction $(F_{(12,520)}=2.41, p<0.01)$. The pattern of results is evident in the summarized AUC

data shown in Fig. 3B: OVX mice were indistinguishable from males (DEX enhanced analgesia in both cases), which were both different from either sham-treated or OVX females given estradiol replacement. Although the specific comparison between sham females receiving either DEX or saline was not significant, there was a tendency (p=0.06, see # in Fig. 3B) for DEX to reduce analgesia in these intact females.

3. Discussion

These data indicate that ovarian hormones modulate the effect of dextromethorphan on morphine analgesia. Enhancement of morphine analgesia occurred in male mice following co-administration with DEX, but the opposite was generally true in females. These differences are probably mediated by estrogen or its metabolite(s) because ovariectomized (OVX) females, like males, showed a robust DEX-mediated enhancement of morphine analgesia. Moreover,



Fig. 3. This figure illustrates the effect of saline or 100 μ g i.c.v. DEX, followed by 5 mg/kg i.p. morphine on TW latencies in either intact males, intact (shamtreated) females, ovariectomized (OVX) females or OVX females with estradiol replacement. Both males and OVX females given DEX and morphine exhibited a significant increase in duration and amount of antinociception as compared to intact females or OVX females receiving estradiol treatment (Panel A). Panel B summarizes these data by showing the area under the analgesic curves (AUC) for each group. Asterisks signify significant differences between DEX- and saline-treated subjects within each sex condition, while the # sign reflects a non-significant (p=0.06) tendency for DEX attenuation of analgesia in the sham surgery group.

OVX mice receiving estradiol replacement also failed to exhibit DEX-enhanced morphine analgesia.

In order to evaluate the influence of DEX directly and to avoid possible sex differences in the peripheral metabolism of DEX (e.g., Ramachander et al., 1978: Labbe et al., 2000), central (i.c.v.) administration was used. There are two other studies reporting DEX effects on opioid analgesia interacting with sex (Holtman et al., 2003; Nemmani et al., 2004), but both of these evaluated the effects of NMDA antagonists delivered systemically. This procedural difference may account for conflicting results across experiments. In our previous report (Nemmani et al., 2004), males showed effects of DEX akin to those we found here, but females did not show any alteration in morphine analgesia by systemic (oral) administration. Following intraperitoneal administration, Holtman et al. (2003) found slightly more DEX-mediated enhancement of morphine analgesia in female than male rats, a discrepancy that may be due to species influences. Unfortunately, there are so few studies that include both male and female subjects that general conclusions about the mechanisms underlying sex-specific modulation of NMDA-opioid interactions are not possible at this time. We would predict that cyclic changes in ovarian hormones might influence the way that DEX interacts with morphine, and also may explain why the attenuation of morphine analgesia by DEX in females was variable (clearly present in Experiment 1, nearly evident in Experiment 2 and absent in Experiment 3) though this hypothesis needs to be tested directly.

Clinical trials using NMDA antagonists (Mercadante et al., 2000; Wu et al., 2000a,b; Heiskanen et al., 2002) have also found enhancement of opiate effects and, as a matter of regulatory requirement, do include both sexes. However, the average age in each of these studies is between 50 and 60 years (i.e., post-menopausal). Moreover, the authors neither analyze the data by sex or age, so it is impossible to discern the relative efficacy in females or how this might depend upon hormonal status. However, Bell (1999) evaluated the effect of adding ketamine to the treatment regimen of opiate tolerant humans in case studies of 3 subjects: 2 males and 1 female. Both males showed dramatic improvement in analgesia levels when morphine was combined with ketamine, but Bell reports that in the female subject analgesia was "insufficient".

Multiple mechanisms no doubt underlie the behavioral and neural modifications associated with chronic opiate use. How precisely NMDA-receptors influence morphine effects is not entirely understood, but over the past several years a dominant model has emerged that suggests opioid potentiation and reversal of tolerance by NMDA receptor antagonists may be due to the same cellular mechanisms (prevention of opioid receptor desensitization; see Trujillo, 2002 for a review). For example, NMDA receptor activation contributes to acute tolerance following a single exposure to an opiate (Larcher et al., 1998) and a lack of acute opioid tolerance (see Wang and Ho, 1994 for review or cf.: Kornetsky and Bain, 1968) may mediate the potentiation of morphine analgesia seen in the presence of NMDA antagonists. Thus, antagonism of NMDA receptor activity may attenuate opioid receptor desensitization, preventing acute tolerance after a single dose of opioid. This protection against acute opioid tolerance may be manifest in behavioral assays as analgesic potentiation. Again, there is ample evidence that NMDA receptor antagonists alter the development and/or expression of tolerance in male mice (Kolesnikov et al., 1988; Kolesnikov and Pasternak, 1999; Lufty et al., 1999; Popik and Kozela, 1999; Belozertseva et al., 2000; Redwine and Trujillo, 2003) and rats (Mao et al., 1996; McNally and Westbrook, 1998; Houghton et al., 2001; Quartaroli et al., 2001; or see Mao, 1999; Trujillo, 2000 for reviews). NMDA receptor antagonists have also been shown to block opiate withdrawal in males (see Mao, 1999; Trujillo, 2000 for reviews). We are currently investigating the ability of NMDA-receptor antagonists to prevent tolerance in females, but for now this is an open question.

The mechanisms underlying estrogen's effect on NMDA receptors have not been fully elucidated, but there is a substantial literature on estrogen modulation of NMDA receptors (see McEwen et al., 1997 for review). For instance, Cyr et al. (2001) review evidence showing that both OVX and estradiol treatment alter NMDA receptor specific binding in various brain regions. Estrogen has also been shown to alter expression of NMDA receptors (D'Souza et al., 2003) and to inhibit both NR1 and NR2a subunit mRNA levels (Gore et al., 2002). In fact, steroids can modulate a variety of voltage and ligandgated ion channels and, in particular, estrogenic compounds directly inhibit NMDA receptor-mediated neuronal responsiveness (Park-Chung et al., 1997; Weaver et al., 1997). Estrogenic steroids can also affect NMDA receptormediated behaviors. For example, Kalkbrenner and Standley (2003) lowered NMDA-induced seizure threshold, as well as hippocampal damage resulting from seizures, in females through ovariectomy. Estrogen also interacts with central NMDA receptors to enhance LTP in the hippocampus (Good et al., 1999) and to provide neuroprotection against antagonist interference with LTP. Gureviciene et al. (2003) report that the blockade of LTP and lowered acquisition of water maze learning by NMDA antagonism, are both ameliorated by estrogen treatment. Taken together with the present results, these studies support a role for estrogen in negative modulation of NMDA receptor activity.

It is worth noting, however, that DEX is not a specific antagonist, and its interaction with opiates could occur through its effects on other systems. For example, nicotinic receptors have been implicated in pain transmission and are also under control of estrogen (Lapchak et al., 1990; Nakazawa and Ohno, 2001; Curtis et al., 2002). In particular, DEX and its metabolite dextrorphan have been found to act as antagonists at the $\alpha_3\beta_4$ neuronal nicotinic receptor (Hernandez et al., 2000; Damaj et al., 2005).

The fact that we saw no evidence of DEX-mediated potentiation of morphine analgesia in intact female subjects suggests that estrogen may mediate qualitative sex differences in the adaptive response to morphine. A better understanding of NMDA-modulation of opiate analgesia and tolerance, as well as the way that these mechanisms interact with sex steroids, may aid in the development of new and better pharmacotherapies to alleviate chronic or severe pain (Wiesenfeld-Hallin, 1998). To help expedite these advances, both basic studies and clinical trials ought to pay particular attention to variables such as sex and hormonal status in order to facilitate understanding of pain sensitivity, antinociception and opiate tolerance in both males and females.

Acknowledgements

We thank Jeffrey S. Mogil for helpful comments on this manuscript.

References

- Advokat C, Rhein FQ. Potentiation of morphine-induced antinociception in acute spinal rats by the NMDA antagonist dextromethorphan. Brain Res 1995;699:157–60.
- Allen RM, Dystra LA. N-Methyl-D-aspartate receptor antagonists potentiate the antinociceptive effects of morphine in squirrel monkeys. J Pharmacol Exp Ther 2001;298:288–97.
- Bell RF. Low-dose subcutaneous ketamine infusion and morphine tolerance. Pain 1999;83:101–3.
- Belozertseva IV, Danysz W, Bespalov AY. Effects of short-acting NMDA receptor antagonist MRZ 2/576 on morphine tolerance development in mice. Naunyn-Schmiedeberg's Arch Pharmacol 2000;361:573-7.
- Ben-Bassat J, Peretz E, Sulman FG. Analgesimetry and ranking of analgesic drugs by the receptacle method. Arch Int Pharmacodyn Ther 1959;122:434–47.
- Bernardi M, Bertolini A, Szczawinska K, Genedani S. Blockade of the polyamine site of NMDA receptors produces antinociception and enhances the effect of morphine in mice. Eur J Pharmacol 1996;298:51–5.
- Bespalov A, Kudryashova M, Zvartau E. Prolongation of morphine analgesia by competitive NMDA receptor antagonist D-Cppene (SDZ EAA 494) in rats. Eur J Pharmacol 1998;351:299–305.
- Bhargava HN. Diversity of agents that modify opioid tolerance, physical dependence, abstinence syndrome, and self administration behavior. Pharmacol Rev 1994;46:293–324.
- Bhargava HN. Enhancement of morphine actions in morphine-naïve and morphine tolerant mice by LY 235959, a competitive antagonist of the NMDA receptor. Gen Pharmacol 1997;28:61–4.
- Boyer JS, Morgan MM, Craft RM. Microinjection of morphine into the rostral bentromedial medulla produces greater antinociception in male compared to female rats. Brain Res 1998;796:315–8.
- Bulka A, Wiesenfeld-Hallin Z, Xu XJ. Differential antinociception by morphine and methadone in two substrains of Sprague–Dawley rats and its potentiation by dextromethorphan. Brain Res 2002;942:95–100.
- Carlezon WA, Kosten TA, Nestler EJ. Behavioral interactions caused by combined administration of morphine and MK-801 in rats. Psycho-pharmacology 2000;151:261-72.

- Chapman V, Dickenson AH. The combination of NMDA antagonism and morphine produces antinociception in the rat dorsal horn. Brain Res 1992;573:321–3.
- Christensen D, Idanpaan-Heikkila JJ, Guilbaud G, Kayser V. The antinociceptive effect of combined systemic administration of morphine and glycine/NMDA receptor antagonist, (+)-HA966 in a rat model of peripheral neuropathy. Br J Pharmacol 1998;125:1641-50.
- Christensen D, Gautron M, Guilbaud G, Kayser V. Combined systemic administration of the glycine/NMDA receptor antagonist, (+)-HA966 and morphine attenuates pain-related behaviour in a rat model of trigeminal neuropathic pain. Pain 1999;83:433–40.
- Cicero TJ, Nock B, Meyer ER. Gender-related differences in the antinoceptive properties of morphine. J Pharmacol Exp Ther 1996;279:767-73.
- Cox BM. Drug tolerance and physical dependence. In: Pratt WB, Taylor P, editors. Principles of Drug Action: The Basis of Pharmacology, 3rd edition. New York: Churchill Livingstone Inc.; 1990. p. 630–90.
- Curtis L, Buisson B, Bertrand S, Bertrand D. Potentiation of human alpha 4 beta 2 neuronal nicotinic acetylcholine receptor by estradiol. Mol Pharmacol 2002;61:127–35.
- Cyr M, Ghribi O, Thibault C, Morissette M, Landry M, Di Paolo T. Ovarian steroids and selective estrogen receptor modulators activity on rat brain NMDA and AMPA receptors. Brain Res Rev 2001;37:153–61.
- Damaj MI, Flood P, Ho KK, May EL, Martin BR. Effect of dextromethorphan and dextrorphan on nicotine and neuronal nicotinic receptors: in vitro and in vivo selectivity. J Pharmacol Exp Ther 2005;312:780–5.
- D'Souza DN, Harlan RE, Garcia MM. Modulation of glutamate receptor expression by gonadal steroid hormones in the rat striatum. Brain Res Bull 2003;59:289–92.
- Good M, Day M, Muir JL. Cyclical changes in endogenous levels of oestrogen modulate the induction of LTD and LTP in the hippocampal CA1 region. Eur J Neurosci 1999;11:4476–80.
- Gore AC, Oung T, Woller MJ. Age-related changes in hypothalamic gonadotropin-releasing hormone and *N*-methyl-D-aspartate receptor gene expression, and their regulation by oestrogen, in the female rat. J Neuroendocrinol 2002;14:300–9.
- Gureviciene I, Puolivali J, Pussinen R, Wang J, Tanila H, Ylinen A. Estrogen treatment alleviates NMDA-antagonist induced hippocampal LTP blockade and cognitive deficits in ovariectomized mice. Neurobiol Learn Mem 2003;79:72–80.
- Heiskanen T, Hartel B, Dahl M-L, Seppala T, Kalso E. Analgesic effects of dextromethorphan and morphine in patients with chronic pain. Pain 2002;96:261–7.
- Hernandez SC, Bertolino M, Xiao Y, Pringle KE, Caruso FS, Kellar KJ. Dextromethorphan and its metabolite dextrorphan block alpha3beta4 neuronal nicotinic receptors. J Pharmacol Exp Ther 2000;293:962–7.
- Holtman Jr JR, Jing X, Wala EP. Sex-related differences in the enhancement of morphine antinociception by NMDA receptor antagonists in rats. Pharmacol Biochem Behav 2003;76:285–93.
- Houghton AK, Parsons CG, Headley PM. Mrz 2/579, a fast kinetic NMDA channel blocker, reduces the development of morphine tolerance in awake rats. Pain 2001;91:201–7.
- Kalkbrenner KA, Standley CA. Estrogen modulation of NMDA-induced seizures in ovariectomized and non-ovariectomized rats. Brain Res 2003;964:244–9.
- Kavaliers M, Galea LA. Sex differences in the expression and antagonism of swim stress-induced analgesia in deer mice vary with the breeding season. Pain 1995;63:324–7.
- Kepler KL, Standifer KM, Paul D, Kest B, Pasternak GW, Bodnar RJ. Gender effects and central opioid analgesia. Pain 1991;45:87–94.
- Kest B, Wilson SG, Mogil JS. Sex differences in supraspinal morphine analgesia are dependent on genotype. J Pharmacol Exp Ther 1999;289:1370-5.
- Kolesnikov Y, Jain S, Wilson R, Pasternak GW. Lack of morphine and enkephalin tolerance in 129/SvEv mice: evidence for a NMDA receptor defect. J Pharmacol Exp Ther 1988;284:455–9.

- Kolesnikov Y, Pasternak GW. Topical opioids in mice: analgesia and reversal of tolerance by a topical *N*-methyl-D-aspartate antagonist. J Pharmacol Exp Ther 1999;290:247–52.
- Kornetsky C, Bain G. Morphine: single-dose tolerance. Science 1968;162:1011-2.
- Kozela E, Danysz W, Popik P. Uncompetitive NMDA receptor antagonist potentiate morphine antinociception recorded from the tail but not the hind paw in rats. Eur J Pharmacol 2001;423:17–26.
- Labbe L, Sirois C, Pilote S, Arseneault M, Robitaille NM, Turgeon J, et al. Effect of gender, sex hormones, time variables and physiological urinary pH on apparent CYP2D6 activity as assessed by metabolic ratios of marker substrates. Pharmacogenetics 2000;10:425–38.
- Lapchak PA, Araujo DM, Quirion R, Beaudet A. Chronic estradiol treatment alters central cholinergic function in the female rat: effect on choline acetyltransferase activity, acetylcholine content, and nicotinic autoreceptor function. Brain Res 1990;20:249–55.
- Larcher A, Laulin JP, Celerier E, Le Moal M, Simonnet G. Acute tolerance associated with a single opiate administration: involvement of *N*-methyl D-aspartate-dependent pain facilitatory systems. Neuroscience 1998;84:583–9.
- Laulin J-P, Maurette P, Corcuff J-B, Rivat C, Chauvin M, Simonnet G. The role of ketamine in preventing fentanyl-induced hyperalgesia and subsequent acute morphine tolerance. Anesth Analg 2002;94: 1263–9.
- Laursen SE, Belknap JK. Intracerebroventicular injections in mice. Some methodological refinements. J Pharmacol Methods 1986;16:355-7.
- Lufty K, Doan P, Weber E. ACEA-1328, and NMDA receptor/glycine site antagonist, acutely potentiates antinociception and chronically attenuates tolerance induced by morphine. Pharmacol Res 1999;40:435–42.
- Mao J. NMDA and opioid receptors: their interactions in antinociception, tolerance and neuroplasticity. Brain Res Rev 1999;30:289-304.
- Mao J, Price DD, Caruso F, Mayer DJ. Oral administration of dextromethorphan prevents the development of morphine tolerance and dependence in rats. Pain 1996;67:361–8.
- McEwen BS, Alves SE, Bulloch K, Weiland NG. Ovarian steroids and the brain: implications for cognition and aging. Neurology 1997;48: S8-15.
- McNally GP, Westbrook RF. Test type influences the expression of lithium chloride-induced hyperalgesia. Pharmacol Biochem Behav 1998;61: 385–94.
- Mercadante S, Arcuri E, Tirelli W, Casuccio A. Analgesic effect of intravenous ketamine in cancer patients on morphine therapy: a randomized, controlled, double-blind, crossover, double-dose study. J Pain Symptom Manage 2000;20:246–52.
- Mogil JS, Sternberg WF, Kest B, Marek P, Liebeskind JC. Sex differences in the antagonism of swim stress-induced analgesia: effects of gonadectomy and estrogen replacement. Pain 1993;53:17–25.
- Nakazawa K, Ohno Y. Modulation by estrogens and xenoestrogens of recombinant human neuronal nicotine receptors. Eur J Pharmacol 2001;430:175-83.
- Nemmani KV, Grisel JE, Stowe JR, Smith-Carliss R, Mogil JS. Modulation of morphine analgesia by site-specific *N*-methyl-D-aspartate receptor antagonists: dependence on sex, site of antagonism, morphine dose, and time. Pain 2004;109:274–83.

- Nishiyama T. Interaction among NMDA receptor-, NMDA glycine site- and AMPA receptor antagonists in spinally mediated analgesia. Can J Anaesth 2000;47:693–8.
- Park-Chung M, Wu FS, Purdy RH, Malayev AA, Gibbs TT, Farb DH. Distinct sites for inverse modulation of *N*-methyl-D-aspartate receptors by sulfated steroids. Mol Pharmacol 1997;52:1113–23.
- Plesan A, Hoffman O, Xu XJ, Wisenfield-Hallin Z. Genetic differences in the antinociceptive effects of morphine and its potentiaton by dextromethorphan in rats. Neurosci Lett 1999;263:53-6.
- Popik P, Kozela E. Clinically available NMDA antagonist, memantine, attenuates tolerance to analgesic effects of morphine in a mouse tail flick test. Pol J Pharmacol 1999;51:223–31.
- Quartaroli M, Fasdelli N, Bettelini L, Maraia G, Corsi M. GV196771A, an NMDA receptor/glycine site antagonist, attenuates mechanical allodynia in neuropathic rats and reduces tolerance induced by morphine in mice. Eur J Pharmacol 2001;403:219–27.
- Ramachander G, Bapatla KR, Emele JF. Sex differences in plasma half-life of dextrorphan in rats administered dextromethorphan. J Pharm Sci 1978;67:1326–7.
- Redwine KE, Trujillo KA. Effects of NMDA receptor antagonists on acute mu-opioid analgesia in the rat. Pharmacol Biochem Behav 2003;76:361-72.
- Sarton E, Olofsen F, Romberg R, den Hartigh J, Kest B, Nieuwenhuiis D. Sex differences in morphine analgesia: an experimental study in healthy volunteers. Anesthesiology 2000;93:1245–54.
- Tiseo PJ, Inturrisisi CE. Attenuation and reversal of morphine tolerance by the competitive *N*-methyl-D-aspartate receptor antagonist, LY274614. J Pharmacol Exp Ther 1993;264:1090–6.
- Trujillo KA. Are NMDA receptors involved in opiate-induced neural and behavioral plasticity? A review of preclinical studies. Psychopharmacology 2000;151:121–41.
- Trujillo KA. The neurobiology of opiate tolerance, dependence and sensitization: mechanisms of NMDA receptor-dependent synaptic plasticity. Neurotox Res 2002;4:373–91.
- Trujillo KA, Akil H. Inhibition of morphine tolerance and dependence by the NMDA receptor antagonist MK-801. Science 1991;251:85–7.
- Wang JJ, Ho ST. Acute and chronic opioid tolerance: a pharmacological review. Acta Anaesthesiol Sin 1994;42:261–7.
- Weaver Jr CE, Marek P, Park-Chung M, Tam SW, Farb DH. Neuroprotective activity of a new class of steroidal inhibitors of the *N*-methyl-D-aspartate receptor. Proc Natl Acad Sci U S A 1997;94:10450–4.
- Wen ZH, Chang YC, Cherng CH, Wang JJ, Tao PL, Wong CS. Increasing of intrathecal CSF excitatory amino acids concentration following challenge in morphine-tolerant rats. Brain Res 2004;995:253–9.
- Wiesenfeld-Hallin Z. Combined opioid-NMDA antagonist therapies: what advantages do they offer for the control of pain syndromes? Drugs 1998;55:1–4.
- Wu CT, Yeh CC, Yu JC, Lee MMS, Tao PL, Ho ST, et al. Pre-incisional epidural ketamine, morphine and bupivacaine combined with epidural and general anaesthesia provides pre-emptive analgesia for upper abdominal surgery. Acta Anaesthesiol Scand 2000a;44:63–8.
- Wu CT, Yu JC, Liu ST, Yeh CC, Li CY, Wong CS. Preincisional dextromethorphan treatment for post operative pain management after upper abdominal surgery. World J Surg 2000b;24:512–7.